

TruSight® RNA Pan-Cancer Panel

Comprehensive assessment of cancer-related RNA transcripts and fusion detection in FFPE tissues and other oncology samples.

Highlights

- Comprehensive View of Cancer Pathways**
 Illumina RNA enrichment chemistry enables interrogation of gene expression levels, variants, and gene fusions
- Low-input Protocol for All Sample Types**
 Optimized for sequencing RNA from all sample types, including formalin-fixed, paraffin-embedded tissues
- Focus on Relevant Transcripts and Fusion Genes**
 Industry validated content for a comprehensive view of 1,385 cancer-related RNA transcripts
- Economical, Targeted RNA-Seq on a Desktop Sequencer**
 Enables RNA sequencing on the MiniSeq™ or MiSeq® Systems for affordable, sensitive analysis

allowing cost effective access to NGS for any lab. TruSight RNA Pan-Cancer Panel can be used for gene expression profiling and variant calling, highlighting functionally relevant mutations. The panel also detects fusion genes that most DNA panels would not detect. The TruSight RNA Pan-Cancer Panel provides a comprehensive assessment of cancer-related RNA transcripts for a more detailed view of cancer biology.

Table 1: Coverage Details

Parameter	Specification
Number of target genes	1,385
Targeted exonic regions	21,043
Number of probes	57,010

Introduction

To date, at least 138 known driver genes for cancer have been discovered.¹ However, large numbers of variants are often detected in DNA of unexpressed genes, highlighting that RNA expression can provide key insights into the functionally relevant genes in cancer. Cancer can arise from epigenetic changes, expression level changes, and gene fusions that are undetectable by standard sequencing.^{2,3} RNA sequencing (RNA-Seq) using next-generation sequencing (NGS) offers the ability to capture all relevant transcripts in a single assay. Compared to traditional array-based approaches, targeted RNA-Seq is a highly sensitive approach, with a broader dynamic range, that robustly detects RNAs of low abundance. In addition, RNA-Seq can identify gene fusions from both known and novel fusion gene partners.

To help clinical researchers gain a deeper understanding of the gene expression patterns in cancer classification and progression, the TruSight RNA Pan-Cancer Panel offers in-depth assessment of cancer-related RNA transcripts, including measurement of expression levels and detection of fusion genes. The panel includes 1,385 genes that have been cited in public databases and implicated in cancer, including solid tumors, soft tissue cancers, and hematological malignancies. TruSight RNA Pan-Cancer Panel enables cost-effective analysis of limited and degraded samples, such as formalin-fixed, paraffin-embedded (FFPE) tumor tissue, on a desktop sequencer.

Comprehensive Coverage of Relevant Genes

The content of the TruSight RNA Pan-Cancer Panel represents 1385 genes implicated in cancer pathways (Table 1). In a single assay, researchers can assess all relevant RNA transcripts for multiple cancer types, regardless of origin. The focus on a subset of relevant genes enables RNA-Seq with high sensitivity on a desktop sequencer,

Efficient Analysis of Difficult Samples

Archival FFPE tissues provide a valuable repository of information for cancer research, but the RNA preserved within these samples is often highly degraded. This degradation poses a challenge when creating libraries for NGS. To overcome these challenges, the TruSight RNA Pan-Cancer Panel is optimized for high performance from both high and low quality RNA sample types, such as bone marrow or FFPE tumor tissue. Libraries can be prepared from as little as 10 ng total RNA, or 20 ng FFPE RNA. This low input requirement makes the TruSight RNA Pan-Cancer Panel ideal for reliable targeted analysis of limited samples (Figure 1).

Simple, Scalable Workflow

The TruSight RNA Pan-Cancer workflow first creates RNA-Seq libraries. This method adds unique oligonucleotide indexes to each library, tagging them for downstream multiplexed sequencing (Figure 2A). Libraries are hybridized to biotin-labeled probes specific for targeted RNA regions (Figure 2B). These targets are then captured by adding streptavidin beads that bind to the biotinylated probes (Figure 2C). Magnets pull the bound fragments from solution (Figure 2D). Captured fragments are eluted from the beads and a second hybridization and elution are performed. After amplification, a targeted library is ready for cluster generation and sequencing.



